



# International Journal of Pharmaceutical Research and Development (IJPRD)

Platform for Pharmaceutical Researches & Ideas

www.ijprd.com

## EFFECT OF BETLE LEAF STALK EXTRACTION ON TESTICULAR METABOLISM IN ALBINO RATS

Changamma C<sup>\*1</sup>,  
Vengaiyah V<sup>1</sup>, Govardhan Naik A<sup>1</sup>, Lalithamma A<sup>1</sup>

<sup>1</sup>Department of Zoology, S.V. University, Tirupati-517 502, A.P, India

### ABSTRACT

*This study aimed to evaluate the betle leaf stalk extraction as an antispermatogenic in male albino rats. The oral administration of betle leaf stalk extraction resulted in significant increase in body weight and testes weight with no significant changes in TSI. The water content in testes does not show any significant changes while dry matter slightly reduced. These results indicate that there was not much effect on structural composition of testes. The biochemical studies indicate there was significant elevation in total proteins and significant reduction in carbohydrates & lipids. The reduction in total lipid content indicating impaired lipid metabolism in the testis. The elevated protein content suggests the accumulation of proteins, which were not utilized for spermatogenesis in the form of proteins or in the form of enzymes. The accumulation of testicular cholesterol by betle leaf stalk extraction indicates reduced steroidogenesis, which leads to impaired spermatogenesis, thus acting as antispermatogenic agent.*

**KEYWORDS :** Betel leaf stalk, Carbohydrates, Proteins, Lipids, Albino rat etc.

### INTRODUCTION

In recent years, plants are perused over synthetic contraceptive drug because plants are easily available, economic and devoid of harmful side effects [1]. The primary requisite of an antifertility agent for human is that it should be non-toxic, non-teratogenic and should not interfere with the normal metabolic and behavioral process. Further, the method should be reversible [2]. Several plant species have been described as antifertility agents [3]. The oral administration of

seed extracts of Cuminum cyminum, fruit extracts of *S. emarginatus*, *T. belerica* and *Allium cepa* were lowered androgen dependent parameters revealing reduction in the circulating androgen [4&5].

The betel plant is an ever green and perennial creeper, with glossy heart shaped and white catkin. The piper betel leaves extract contains large number of bio active molecules like poly phenol, alkaloids, steroids, saponin and tannin. Piper betel has light yellow aromatic

### Correspondence to Author

**Dr C. Changamma**

Department of Zoology, S.V.  
University, Tirupati-517 502, A.P,  
India

**Email:** challa1957@gmail.com

essential oil, with sharp burning taste [6]. Its promising traditional applications have led to many chemical and biological studies. The P. betle treatment caused reduction in reproductive organ weights, as compared to control value. The data suggests that the P. betle ethanolic extract exerted antifertility and antiestrogenic effects in female rats. The effects brought by P. betle extract are non-toxic and transient [1].

A study to develop an orally effective male contraceptive agent was extensively carried out in male mice with various doses of the leaf-stalk extracts of Piper betel. The results show no toxicity in all metabolically active tissues of mice and interestingly, the contraceptive efficacy emphasised reversible fertility after withdrawal of treatment [7]. There was no biochemical studies has been carried out in male rats so far, hence the present investigation was undertaken to focus on antifertility efficacy of Betel leaf stalk ethanolic extract on male albino rats.

## MATERIALS AND METHODS

Healthy adult male Wister strain albino rats (90 day old, weight  $230 \pm 10$ g) were administered with 50 mg/kg body wt/day of alcoholic extract of betel leaf stalk through oral route for 15 days by Gavages method. The alcoholic extract was prepared according to [8] WHO (1983) protocol CG-04. The betel leaf stalk was dried, powdered and extracted with 95% ethanol (v/v) at 55-60°C for 3 h. The solvent was distilled under reduced pressure; the resulting mass was dried under vacuum and kept at 24°C until use. The control animals were given normal saline or sterile distilled water. Both control and experimental rats were maintained under standard animal house facilities, with a temperature of  $25 \pm 2$ °C, and 12h dark & 12 h day light, and fed on standard rat feed obtained from Hindustan Lever Ltd., Mumbai, India.

Twenty four hours after the last dose, the animals were autopsied and testes, epididymis, seminal vesicles, prostate gland and liver were isolated; blood serum was collected by puncturing the heart, chilled immediately and used for biochemical analysis. The TSI (Tissue Somatic Index) dry mater & water content were analyzed

gravimetrically. The total proteins[9], total carbohydrates[10], total lipids[11], total cholesterol[12], free fatty acids[13] and Glycerol[14] were estimated biochemically both in control and experimental rat tissues.

## RESULTS AND DISCUSSION

The data represented in table-1to 3 indicates the gravimetric analysis, proximate analysis and lipid profiles in testes of control and betle leaf stalk extraction treated rats. The oral administration of betle leaf stalk extraction resulted in significant increase in body weight and organ weight with no significant changes in TSI[15]. The water content does not shows any significant changes while dry matter slightly reduced. These results indicate that there was not much effect on structural composition of testes. The biochemical studies indicate there was significant elevation in total proteins and significant reduction in carbohydrates& lipids.

The testis, a primary sex organ is the site for spermatogenesis. The leaf stalk extraction as an antispermatogenic, it shows its effect on spermatozoa with changes in seminal plasma composition.

The proteins, carbohydrates and lipids are the major fuel for mammalian organisms. The carbohydrates are the first fuel and lipids are the second major fuel. The testicular carbohydrates were decreased over control, indicates the impaired carbohydrates, lead to impaired spermatogenesis. Generally the reproductive tissues like testes largely depend on the carbohydrates for spermatogenesis [16&17]. The mature germ cell population depends on testicular carbohydrate reserves, suggesting impaired germ cell structure and function of spermatocytes, spermatids and spermatozoa [18&19]. Hence the reduced carbohydrates by leaf stalk extraction represents the reduced spermatogenesis thus shows its antispermatogenic effect.

Apart from carbohydrates, lipids are the second major fuel for mammalian organisms. Lipids are water-insoluble biomolecules and have a

variety of biological roles: as energy stores and fuel molecules and as signal molecules and structural components of membranes [20]. There was significant decrease in total lipid content indicating impaired lipid metabolism in the testis [21]. The elevated protein content suggests the accumulation of proteins, which were not utilized for spermatogenesis in the form of proteins or in the form of enzymes.

The cholesterol levels were significantly increased in testes. Cholesterol is the precursor of the steroid hormones [22], providing the backbone of the steroid molecule. The biosynthesis of testosterone directly from cholesterol can only occur in the Leydig cells [23]. Furthermore, it is the precursor molecule of steroid hormones, such as progesterone, testosterone and cortisol. The unsaponifiable fraction of the neutral lipid fraction of the rat testis represents primarily cholesterol and steroids [24]. Cholesterol is an important precursor for the steroid hormones. The testis and its metabolism are dependent on the plasma and endogenously synthesized cholesterol. Hence the

cholesterol levels were estimated and found to be significantly increased in the experimental rat testis. This observation indicates either its increased uptake from the plasma or increased synthesis or decreased mobilization towards androgenesis or decreased catabolism.

The accumulation of testicular cholesterol by betle leaf stalk extraction indicates reduced steroidogenesis, which leads to impaired spermatogenesis, thus acting as antispermatogenic agent [25]. The treatment does not show any effect on free fatty acids. The free fatty acid (FFA) is primary fuel for skeletal muscle during sub maximal work [26]. Hence, there was no effect of betel leaf stalk extraction on skeletal muscle. However the glycerol concentration was elevated. Thus it is concluded that the betle leaf stalk extraction shows antispermatogenic effect.

#### ACKNOWLEDGMENT

The authors are grateful to BSR, RGNF, MANF-UGC, New Delhi, for financial assistance.

**Table: 1** Effect of Betel leaf stalk extraction on **Body weight, Organ weight, TSI, Dry matter & Water content** in **testes** over Control rats.

S.No	Parameter	Control	Betel leaf stalk extract treated	% Change
1	Body weight(g)	236.21± 21.47	265.31±23.74	+12.29 ***
2	Organ Weight( g)	2.54±0.12	2.78±0.11	+9.44 **
3	TSI	1.07±0.10	1.04±0.12	-3.25 NS
4	Dry matter (mg/g wet wt.)	305.31 ± 28.43	272.73 ± 24.18	-10.66 ***
5	Water content (mg/g wet wt.)	732.56 ± 70.63	763.44± 75.45	+4.21 NS

Mean± SD of six individual observations. + And – indicates percent increase and decrease respectively over control. \*\*indicates P<0.01, \*\*\*indicates P<0.05, NS indicates non significant changes.

**Table 2:** Effect of Betel leaf stalk extract on **Total Proteins, Total Carbohydrates** and **Total Lipids** in **testis** over control rats.

S.NO	Parameter	Control	Betel leaf stalk extract treated	% Change
1	Total Proteins (mg/g wet weight)	164.21 ± 10.73	198.16 ± 12.43	+20.67*
2	Total Carbohydrate (mg/g wet weight)	35.34 ± 1.14	28.79 ± 1.32	-18.53*
3	Total Lipids (mg/g wet weight)	80.10 ± 6.32	65.81 ± 3.42	-17.84*

Mean± SD of six individual observations. + And – indicates percent increase and decrease respectively over control. \* indicates P<0.001 the level of significance, NS indicates non significant changes.

**Table 3:** Effect of Betel leaf stalk extraction on **Cholesterol, Free Fatty acids** and **Glycerol** in **Testes** over control rats.

S.NO	Parameter	Control	Betel leaf stalk extract treated	% Change
1	Cholesterol (mg/g wet wt.)	2.01 ±0.12	3.19 ±0.23	+59.71*
2	Free Fatty Acids (mg/g wet wt.)	21.60 ±1.82	22.16 ±1.74	+2.59 NS
3	Glycerol (mg/g wet wt.)	18.83±1.23	36.67±2.01	+94.74*

Mean± SD of six individual observations. + And – indicates percent increase and decrease respectively over control. \* indicates P<0.001 the level of significance, NS indicates non significant changes

## REFERENCES

- Sharma JD, Lalita Sharma and Poonam yadav, Antifertility Efficacy of Piper betle Linn. (Petiole) on Female Albino Rats, Asian J. Exp. Sci., Vol.21(1), 2007,145-150.
- Ashu Chaudhary N, Sharma P, Sharma , Jasuja ND, Sharma G, Joshi SC and Singh RV, Contraceptive property of nitrogen / oxygen sulfur donor heterocyclic compounds, RJC Rasayan J. Chem Vol.1(3), 2008, 648-692.
- Lin CC, Crude drugs used for the treatment of diabetes mellitus in Taiwan, Am J Clin Med 20, 1992, 269-279.
- Priya G, Saravanan K. and Renuka C. Medicinal plants with potential antifertility activity- A review of sixteen years of herbal medicine research (1994-2010) International Journal of Pharm Tech Research Vol.4(1), 2012, 481-494.
- Venkatesh V, Sharma JD. and Raka Kamal, A comparative study of effect of Alcoholic Extract of Sapindus emarginatus, Terminalia belerica,

- Cuminum cyminum and Allium cepa on Reproductive organs of male Albino rats. Asian.J. Exp. Sci, 16(1&2), 2002, 51-63.
6. Chandra Vikash, Tripathi Shalini, Verma NK, Sing DP, Chaudhary SK, Roshan Asha, Piper betel: Phyto Chemistry, Traditional use & Pharmacological activity- A Review, International Journal of pharmaceutical Research and Development, IJPRD, 2011; Vol 4 (4): 2012, 216 - 223.
  7. Sarkar M, Gangopadhyay P, Basak B, Chakrabarty K, Banarji B, Adikary P, and Chatterjee A, The reversible antifertility effect of Piper betel Linn. On Swiss albino male mice. Contraception. 62(5): 2000, 271-274.
  8. WHO: Protocol CG-40 (1983) Preparation of Alcoholic Extract for Bioassay and Phytochemical Studies. (APJ F/IP, 1001A
  9. Lowry OH, Rosebrough NJ, Farr AL, Randall RJ Protein measurement with the Folin phenol reagent. J Biol Chem. 193: 1951, 265-270.
  10. Carrol N V, Longley H M and Roe J H, Glycogen determination in liver and muscle by use of anthrone reagent. J. Biol Chem 220:1956, 583-95.
  11. Folch JM, Lees MP and Stana-stanley GH, A simple method for the isolation and purification of total lipids from animal tissues. J Biol Chem 226:1957, 497-505.
  12. Natelson, S, Total cholesterol procedure Liebermann-Burchard reagent. In: techniques of clinical chemistry, Charles, C. Thomas Publishers, Springfield, Illinois, USA, 3<sup>rd</sup> edn. 1971, 263-270.
  13. Natelson S, In techniques of chemical Chemistry (ed) C T Charles, Thomas (1997) Publishers, USA, 1965.
  14. Burton RN, Methods in Enzymology, 1957, 246-248
  15. Sathiyaraj K, Sivaraj A, Madhumitha G, Vinoth kumar P, Mary saral A, Devi K, Senthil Kumar B, Antifertility effect of aqueous leaf extract of aegle marmelos on male albino rats International Journal of Current Pharmaceutical Research, Vol 2(1), 2010, 26-29.
  16. Leiderman B & Mancini, RE, Glycogen content in the rat testis from postnatal to adult Ages . Endocrinology, 95: 1969, 607-612.
  17. Ewing, IL. Means, AF, Beams, CG, Montgomery, JR & Montgomery, RD, Biochemical changes in rat testes during postnatal maturation. J Reprod Fertil., 48:1966,265-270.
  18. Udoh FV, Uodh PB and Umoh EE, Activity of Alkaloid extract of Carica papaya seeds on Reproductive functions in male Wistar rats. Pharmaceutical Biology, 4 (3), 2005,563-567
  19. Lohiya NK, Mishra PK, Pathak N, Manivannan B, Bhande S, Panneerdoss S and Sriram S, Efficacy trial on the purified compounds of the seeds of Carica papaya for male contraception in albino rat. Reproductive Toxicology, 20(1): 2005, 135-48.
  20. Michael M.Cox and David L. Nelson. Lehninger Principles of Biochemistry, V Edition, 2008, 371-418.
  21. Changamma C, Lakshman J, Hasim Basha S, and Govardhan Naik A, Effect of 5Thio-D Glucose on Testicular Metabolism of Albino Rat, Journal of Applied Sciences Research, 7(2): 2011, 98-101.
  22. Payne A H, Youngblood GL, Regulation of expression of steroidogenic enzymes in Leydig cells. Biol Reprod, 52: 1995, 217–225.
  23. Jurgens MH, Blunn CT and Peo ER, JR. Vitamin D2 and Cholesterolemia in the Growing Rat J. Nutr., 101: 1971, 153- 160.
  24. Litscher ES, Williams Z, Wassarman PM, Zona pellucid glycoprotein ZP3 and fertilization in Mammals. Mol Reprod Dev. 76: 2009, 933–41.
  25. Arvind Singh, Sushila Kala, Deepak N, Kapoor, Richa Gupta, Antar Virk, Samarjeet Singh and Jyoti Chaudhary, Effect on human sperm mitochondrial activity by Piper betle and Calendula officinalis, Annals of Biological Research, 2 (5) 2011, 622-627
  26. Palmer WK, Caruso RA and Oseai LB, Possible role of lipoprotein lipase in the regulation of endogenous triacylglycerols in the rat heart. Biochem.J. 198: 1981, 159-166.

\*\*\*\*\*